

PRODUCT INFORMATION SHEET

Catalog number: 21027 Unit size: 10x50 ug

Fura-FF, AM [Fura-2FF, AM] *CAS 348079-12-9*

Component	Storage	Amount
Fura-FF, AM [Fura-2FF, AM] *CAS 348079-12-9*	Freeze (< -15 °C), Minimize light exposure	10x50 ug

OVERVIEW

Among the ratiometric calcium indicators, Fura-2 and Indo-1 are most commonly used. Fura-2 is excitation-ratioable while Indo-1 is emission-ratioable. Fura-2 is preferred for ratio-imaging microscopy, in which it is more practical to change excitation wavelengths than emission wavelengths. Upon binding Ca2+, Fura-2 exhibits an absorption shift that can be observed by scanning the excitation spectrum between 300 and 400 nm, while monitoring the emission at ~510 nm. The cell-permeant Fura-2FF AM is an analog of Fura-2 AM with much lower calcium binding affinity, Kd ~10 μ M. This AM ester form can be loaded into live cells noninvasively.

KEY PARAMETERS

Fluorescence microscope

Excitation Emission Recommended plate

Fluorescence microplate reader

Fura 2 filter set Fura 2 filter set Black wall/clear bottom

340, 380 510 475 Black wall/clear bottom Bottom read mode/Programmable liquid handling

PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

Fura-FF AM Stock Solution

Prepare a 2 to 5 mM stock solution of Fura-FF AM in high-quality, anhydrous DMSO.

PREPARATION OF WORKING SOLUTION

Fura-FF AM Working Solution

On the day of the experiment, either dissolve Fura-FF AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature. Prepare a dye working solution of 2 to 20 μ M in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Fura-FF AM at a final concentration of 4-5 μ M is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

Note The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Fura-FF AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

Note If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of ReadiUse[™] probenecid products, including water-soluble, sodium salt, and stabilized solution, can be purchased from AAT Bioquest.

SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

- 1. Prepare cells in growth medium overnight.
- On the next day, add 1X Fura-FF AM working solution into your cell plate.

Note If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

Note Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.

- Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
- Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a Fura 2 filter set or a fluorescence plate reader containing a programmable liquid handling system such as a FlexStation, at Ex/Em 1 = 340/510 nm cutoff 475 nm and Ex/Em 2 = 380/510 nm cutoff 475 nm.

EXAMPLE DATA ANALYSIS AND FIGURES

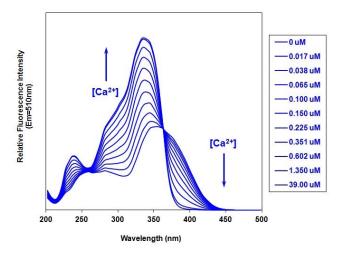


Figure 1. Fluorescence excitation spectra of Fura-2TM in the presence of 0 to 39 μM free Ca2+.

DISCLAIMER

AAT Bioquest provides high-quality reagents and materials for research use only. For proper handling of potentially hazardous chemicals, please consult the Safety Data Sheet (SDS) provided for the product. Chemical analysis and/or Tel: 408-733-1055 LFax: 408-733-1304 L Email: support@aathio.com

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