

# Rhod-4™. AM

Catalog number: 21120, 21121, 21122, 21123 Unit size: 1 mg, 5x50 ug, 10x50 ug, 20x50 ug

Component	Storage	Amount (Cat No. 21120)	Amount (Cat No. 21121)	Amount (Cat No. 21122)	Amount (Cat No. 21123)
Rhod-4™, AM	Freeze (< -15 °C), Minimize light	1 vial (1 mg)	5x50 ug	10x50 ug	20x50 ug
	exposure				

### **OVERVIEW**

Calcium measurement is critical for numerous biological investigations. Fluorescent probes that show spectral responses upon binding Ca2+ have enabled researchers to investigate changes in intracellular free Ca2+ concentrations by using fluorescence microscopy, flow cytometry, fluorescence spectroscopy and fluorescence microplate readers. Rhod-2 is most commonly used among the red fluorescent calcium indicators. However, Rhod-2 AM is only moderately fluorescent in live cells upon esterase hydrolysis, and has very small cellular calcium responses. Rhod-4 $^{\rm TM}$  has been developed to improve Rhod-2 cell loading and calcium response while maintaining the spectral wavelength of Rhod-2. In CHO and HEK cells Rhod-4 $^{\rm TM}$  AM has cellular calcium response that is 10 times more sensitive than Rhod-2 AM. AAT Bioquest offers versatile packing sizes of Quest Rhod-4 to meet your special needs, e.g., 1 mg; 10x50 μg; 20x50 μg; HTS packages with no additional packaging charge.

#### **KEY PARAMETERS**

### Fluorescence microscope

Excitation TRITC filter set
Emission TRITC filter set
Recommended plate Black wall/clear bottom

### Fluorescence microplate reader

Excitation 540 Emission 590 Cutoff 570

Recommended plate Black wall/clear bottom

Instrument specification(s)

Bottom read mode/Programmable liquid

handling

## PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

## Rhod-4™ AM Stock Solution

Prepare a 2 to 5 mM stock solution of Rhod-4™ AM in high-quality, anhydrous DMSO.

### PREPARATION OF WORKING SOLUTION

## Rhod-4™ AM Working Solution

On the day of the experiment, either dissolve Rhod-4 $^{\text{TM}}$  AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature. Prepare a dye working solution of 2 to 20  $\mu$ M in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Rhod-4 $^{\text{TM}}$  AM at a final concentration of 4-5  $\mu$ M is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

Note The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Rhod-4™ AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

**Note** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of ReadiUse<sup>™</sup>

probenecid products, including water-soluble, sodium salt, and stabilized solution, can be purchased from AAT Bioquest.

### SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

- 1. Prepare cells in growth medium overnight.
- On the next day, add 1X Rhod-4™ AM working solution into your cell plate.

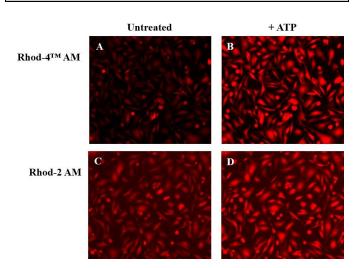
**Note** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

 Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

**Note** Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.

- Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
- 5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a TRITC filter set or a fluorescence plate reader containing a programmable liquid handling system such as an FDSS, FLIPR, or FlexStation, at Ex/Em = 540/590 nm cutoff 570 nm.

## **EXAMPLE DATA ANALYSIS AND FIGURES**



**Figure 1.** ATP-stimulated calcium responses of endogenous P2Y receptors were measured in CHO-K1 cells with Rhod-4<sup>™</sup> AM (Cat# 21120) and Rhod-2 AM (Cat# 21064). CHO-K1 cells were seeded overnight at 50,000 cells/100 μL/well in a Costar 96-well black wall/clear bottom plate. The growth medium was removed, and the cells were incubated with 100 μL of dye loading solution using Rhod-4<sup>™</sup>

AM (4  $\mu$ M, A and B) or Rhod-2 AM (4  $\mu$ M, C and D) for 1 hour in a 37 °C, 5% CO2 incubator. The staining solution was replaced with 200  $\mu$ L HHBS, then the cells were imaged before (A and C) and after (B and D) ATP treatment with a fluorescence microscope (Olympus IX71) using TRITC channel.

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